FIGURE 1

Replace

with

Minicell purification procedure without the inventive techniques

Overnight culture of minicell producing bacterial cells

Differential centrifugation @ 2000g 20 min Pellet most bacterial cells while minicells remain In supernatant

First Cross-flow filtration through 0.45 µm filter Reduce parent bacterial cell contamination minicells are 0.4um diameter & filter through but Some loss of minicells

Second Cross-flow filtration through 0.45 µm filter Further reduce parent bacterial cell contamination. Some loss of minicells.

Cross-flow filtration through 0.2 µm filter Retain minicells but eliminate all contaminants smaller than 0.2um in size

Cross-flow filtration through 0.1μm filter Concentrate minicells

Dead-end filtration through 0.45µm filter Eliminate most of the residual parent bacterial cells

Concentrate minicells through 100kDa filter

Inventive steps described in this patent application

2x Density gradient centrifugation using isotonic and non-toxic density gradient media e.g. OptiPrep.

Removes most parent bacterial cells with minimal loss of minicells

Cross-flow filtration through 0.45 um filter Reduces parent bacterial cell contamination

Stress-induced filamentation of residual parent bacterial cells e.g. incubate minicell suspension with growth media and 5% NaCl (stress inducer) for 4 hrs. Residual parent bacteria turn into filaments and are blocked in subsequent filtration process

Treat with antibiotic that parent bacteria are sensitive to.

Kills all live bacteria present in the preparation

Cross-flow filtration through 0.2 um filter Eliminates all contaminants smaller than 0.2 um. Dead bacterial cells, filaments and minicells are retained.

Dead-end filtration also eliminates filamentous dead parent bacteria

minicells with endotoxin Incubate removal system e.g. Anti-Lipid A antibody conjugated to magnetic beads. The free lipopolysaccharide 1 Anti-Lipid A conjugated magnetic beads are removed from the suspension by placing the tube in a magnetic stand to immobilise the complex and minicells can be collected. This step ensures that sufficient free endotoxin is removed from the minicell suspension to enable in-vivo use of minicells in humans or animals.

Figure 2

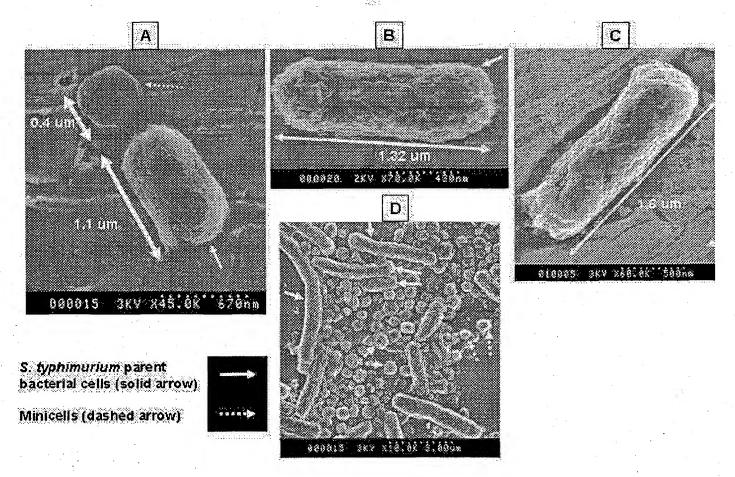
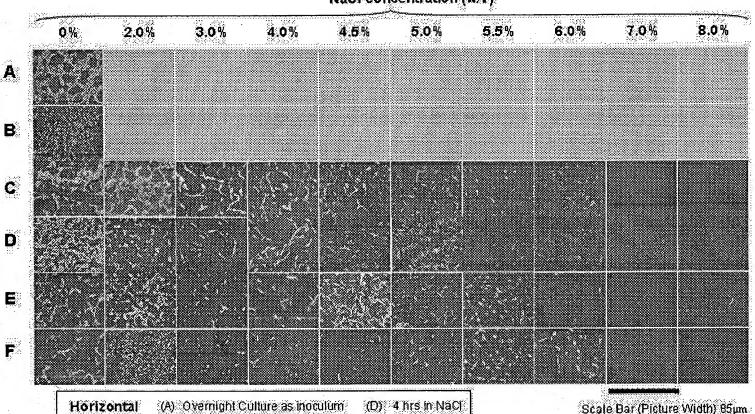


Figure 3 A Filamentation of S. typhimurium [ENSm001] after addition of NaCl (1250x Magnification) NaCI concentration (w/v)



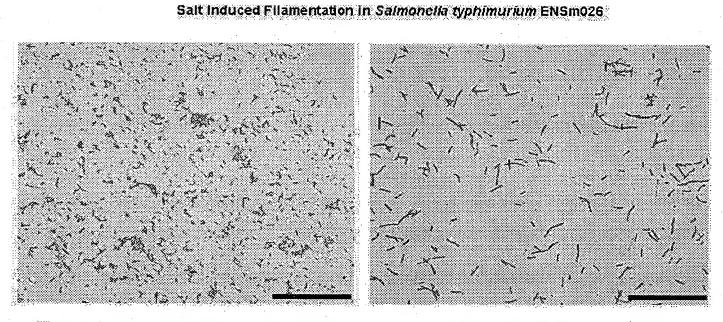
panels

- (A) Overnight Culture as inoculum
 (B) O Hours Time of NaCl Addition
 (C) 2 hrs in NaCl
- (D) 4 hrs in NaCl (E) 8 hrs in NaCl

(F) 24 hrs in NaCl

Scale Bar (Picture Width) 85µm

Figure 3 B



4 hours after Log Culture Inoculum (0% NaCl)

4 hours after Log Culture Inoculum (5.0% NaCl)

Scale Bar 50 µm

Figure 4 A
Filamentation of *E. coli* after addition of NaCl (1250x Magnification)
NaCl concentration (w/v)

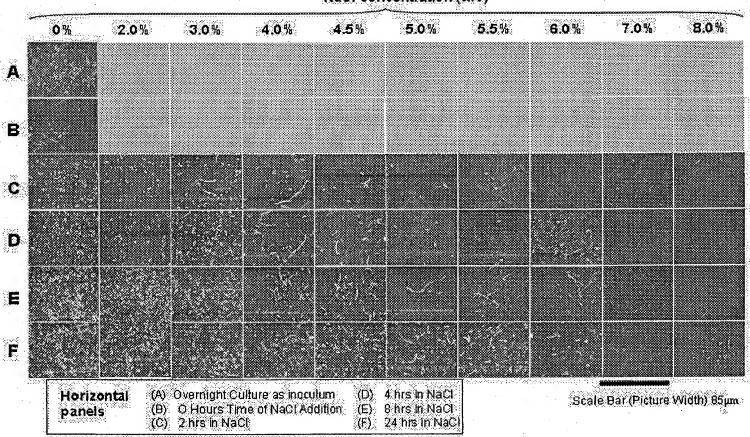
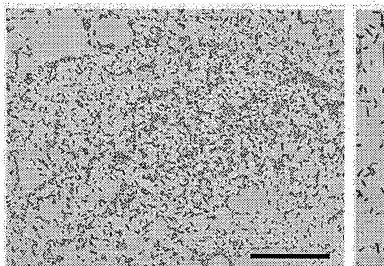
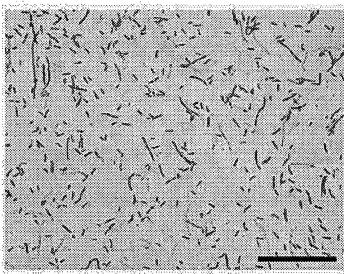


Figure 4 B

Salt Induced filamentation in E. coli



4 hours after Log Culture Inoculum (0% NaCI)



4 hours after Log Culture Inoculum (5.0% NaCl)

Scale Bar 50µm